

Decreased levels of plasma preptin in female patients with knee osteoarthritis

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Abstract

Aim: Osteoarthritis (OA) is a common joint disease which is caused by the effects of mechanical, genetic and biochemical factors. In studies conducted in recent years, it has been shown that osteoblasts play roles in OA pathogenesis. In this study, we investigated the relation between the level of preptin, which is known to have an effect on osteoblast proliferation and differentiation, bone metabolism, and the OA disease.

Materials and Methods: A total of 40 healthy control patients and 40 women who were diagnosed with knee OA were included in the present study. The plasma preptin levels of the individuals who were included in the study were measured with Enzyme-linked Immuno-Sorbent Assay (ELISA) Method. Blood glucose, triglyceride, High-Density Lipoprotein Cholesterol (HDL-c), Low-Density Lipoprotein Cholesterol (LDL-c), total cholesterol levels and Body Mass Indices (BMI) leukocyte, lymphocyte, neutrophil, Erythrocyte Sedimentation Rate (ESR), C-Reactive Protein (CRP) of all participants were measured.

Results: The preptin levels of the knee OA patients in plasma were significantly lower (206 ± 103 ng/L) compared to the healthy control Group (501 ± 525 ng/L) ($p < 0.001$). In addition, the (CRP) levels ($p < 0.02$) and the Body Mass Indices (BMI) ($p < 0.001$) were also higher in the OA Group compared to the Control Group.

Conclusion: In the present study, it was determined that there is a significant relation between knee OA disease and plasma preptin levels. Preptin may have roles in the pathogenesis of the OA disease. Further studies are needed to elucidate the mechanisms through which molecular mechanisms preptin is related to knee OA.

Keywords: Osteoarthritis; knee; preptin; ELISA; plasma

INTRODUCTION

Osteoarthritis (OA) is a common joint disease which is caused by the effects of mechanical, genetic and biochemical factors (1,2). Aging is a major risk factor in OA. In aging, changes occur in the chondrocytes in the joint cartilage and in the destruction of the matrix components (3). Progressive loss of cartilage is accompanied by subchondral bone remodeling, osteophyte formation, and synovial inflammation and pain (4). About 40% of the adult population over the age of 65 years has symptomatic knee or hip OA. Knee OA is more common than hip OA (5,6). Knee OA is one of the reasons for mobility limitation and disability (7). In a study conducted in Turkey, in individuals over 50 years of age with symptomatic knee OA, the

prevalence was found as 14.8%. The prevalence of knee OA was 22.5% in women and 8% in men (8).

Preptin, a peptide consisting of 34 amino acids, was discovered in 2001. Preptin was found to be secreted from isolated pancreatic beta cells with insulin and amylin (9). Although the physiological role of preptin has not been fully elucidated, it has been shown to have an anabolic effect for bone formation (10). Preptin has also been shown to have a proliferative effect in osteoblasts in vitro and to increase bone mineralization in vivo (11). In a study on humans, serum preptin levels were shown to be low in osteoporosis and osteopenia patients (12). In addition, a positive correlation was found between the level of markers of bone production and serum preptin level (13).

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The effect of preptin on bones and its relationship with bone diseases suggest that preptin may also be associated with OA disease. In this study, we investigated the relationship between knee OA disease and plasma preptin level.

MATERIAL and METHODS

Subject

The present study was conducted in accordance with the protocol approved by Bozok University Local Ethics Committee (2017-KAEK-189_2019.01.02_03); and according to the Helsinki Declaration (1975, as revised in 2000). Forty knee OA patient women between the ages of 45 and 82 applying to orthopedics clinic, and 40 healthy controls were included in the study following their informed consents were obtained. The diagnostic criteria (clinical and radiological) were based on American College of Rheumatology Criteria (1986), and on the Kellgren and Lawrence scores (14,15). The patient (study) group consisted of Grade 3 (n=12) and Grade 4 (n=28) patients who had knee plain radiographs and OA diagnosis. The Control Group consisted of those who had knee plain radiographs because of other complaints. Exclusion criteria were having previous knee injury or joint infection, secondary posttraumatic OA, systemic inflammatory or autoimmune disorders, known malignant tumor, end-stage renal or hepatic disease, diabetes, and history of corticosteroid medication. Leukocyte, neutrophil, lymphocyte, C-Reactive Protein (CRP), Erythrocyte Sedimentation Rate (ESR), blood glucose, triglyceride, Low-Density Lipoprotein Cholesterol (LDL-c), High-Density Lipoprotein Cholesterol (HDL-c), total cholesterol levels and Body Mass Indices (BMI) of all participants were measured.

Biochemical Analysis

Blood samples were taken from patients and the control group between 09.00 - 10.00 am; and they were taken to vacutainers with Na₂-EDTA (1.5 mg/mL). The complete blood count, blood glucose level, blood lipid data of the groups were received from hospital registry system. After blood collection, samples were centrifuged at 3000 rpm for 10 min for plasma isolation. The supernatant was removed rapidly and was kept frozen at -80°C until the assays, which were carried out a specialist blind to patients determination of plasma preptin levels.

The Enzyme-linked Immuno-Sorbent Assay (ELISA) kit with a minimum detectable concentration of 10 ng/mL and 4000 ng/mL was employed in determining the plasma preptin levels (Bioassay Technology Laboratory, Shanghai, China). The Spectramax ELISA reader (Molecular Devices) was used to determine the optical density values of the samples and standard samples at 450 nm. The results are presented as ng/L.

Statistical Analysis

The data analysis was carried out with the SPSS 20 Package Program. The normality of the quantitative data was checked with the Shapiro-Wilk normality test; and the Independent-samples t-test was employed to

compare normally-distributed data between groups. The non-normally distributed data were compared by using the Mann-Whitney U test. The correlation analysis of the data with normal distribution was carried out with the Pearson Correlation Analysis; and the correlation analysis of the data not showing normal distribution was carried out with Spearman's Correlation Analysis. To define the quantitative data, arithmetic mean \pm standard deviation (SD) was employed; and $p < 0.05$ was considered to be statistically significant.

RESULTS

The statistical analysis results for all the data are given in Table 1. The plasma preptin levels were low at a significant level in the OA Group (206 \pm 103 ng/L) compared to the Control Group (501 \pm 525 ng/L) ($p < 0.001$). It was determined that the CRP level of the OA Group (4.41 \pm 6.79) was higher at a significant level than the Control Group (2.76 \pm 2.39) ($p < 0.02$). It was also determined that the BMI values were higher at a significant level in the OA Group (32 \pm 5.3) than in the Control Group (29 \pm 3.6) ($p < 0.025$). No correlations were detected between the preptin and OA disease stage and all other parameters.

Table 1. Baseline clinical and laboratory characteristics

Variables	Control (n= 40)	OA (n= 40)	P value
Age	60 \pm 6	63 \pm 9	0.9
BMI (kg/mm ²)	29 \pm 3.6	32 \pm 5.3	0.001*
FBG (mg/dL)	94 \pm 11,9	96 \pm 12.3	0.341
Leukocyte (mm ³)	7.4 \pm 2.21	7.02 \pm 1.9	0.279
Lymphocyte (mm ³)	2.9 \pm 4.2	2.7 \pm 4.2	0.616
Neutrophils (mm ³)	4.36 \pm 1.64	4.18 \pm 1.47	0.705
CRP (mg/dl)	2.76 \pm 2.39	4.41 \pm 6.79	0.02*
TG mmol/L	128 \pm 60	150 \pm 80	0.736
LDL-c mmol/L	118 \pm 39	127 \pm 25	0.300
HDL-c mmol/L	58.1 \pm 14.4	57 \pm 16.2	0.765
TC mmol/L	205 \pm 38	216 \pm 33	0.205
ESR (mm/h)	17 \pm 6.9	20 \pm 12	0.158
Preptin (ng/L)	501 \pm 525	206 \pm 103	0.001*

All values are presented as mean \pm SD. * $P < 0.05$ compared with control group. BMI indicates Body Mass Index; CRP, C-Reactive Protein; ESR, Erythrocyte Sedimentation Rate; FBG, Fasting Glucose; TG, Triglyceride; LDL-c, Low-Density Lipoprotein Cholesterol; HDL-c, High-Density Lipoprotein Cholesterol; TC, Total Cholesterol.

DISCUSSION

As the most common joint disease, OA reduces quality of life significantly by causing movement limitation and disability. Although it is also seen in the hands, hips and the spine, it is mostly seen on the knees. The incidence of OA increases with advanced age, and is

higher in females than in males (7,8). For this reason, we conducted our study in women who were at and over the age of 45 with knee OA disease. It was reported that estrogen deprivation that occurs in elderly women is the basic reason of the destruction of cartilage that causes OA (16). Furthermore, it is also known that some other factors (genetic, obesity and mechanical effects on the joint) contribute to OA risk (17). Some theories have been put forward on OA formation one of which arguing that OA initiates with chondrocyte metabolism and cartilage destruction disorder. According to another evidence, synovitis is the primary triggering mechanism of OA process leading to cartilage damage (18). Moreover, according to recent evidence, subchondral bone might be held responsible for articular cartilage degeneration of the exaggerated bone formation (17,19). There are many studies suggesting that Osteocalcin, which is one of the biomarkers of bone formation, is associated with OA (20). There are many studies suggesting that other biomarkers that have physiological roles in bone formation are also associated with OA (17). It has been shown in previous studies that the substances that affect bone formation also have physiological roles related to regulating the energy metabolism (21).

In 2001, preptin was discovered as a peptide that was derived from proinsulin-like growth factor II (proIGF-II). The presence of proIGF-II, which is known to have mitogenic effect, in serum and many tissues in humans and animals was determined (22,23). Preptin was firstly known for its effect to increase the release of glucose-mediated insulin from the pancreas (9). The number of studies conducted on preptin is very few. In preptin-related studies, the focus is mostly on its relation with energy and bone metabolism (24). It was determined that the treatment of preptin causes osteoblast proliferation and differentiation in vitro, but it also suppresses apoptosis in osteoblasts. In the same study, the anabolic effect of preptin on bone was shown in vivo (11). Osteoblasts and chondrocytes are derived from the same stem cells (pluripotent mesenchymal stem cells) (25,26). With the evidence reported in recent studies, it has been shown that osteoblasts play roles in OA pathogenesis (27). It was shown in previous studies that there is a close relation between nuclear factor-kappaB ligand (RANKL)/RANK/osteoprotegerin (OPG) system and the subchondral bone alteration seen in OA (28,29). It was also reported in some previous studies that abnormal mineralization occurs in the subchondral bone in the pathogenesis of OA (30). In the study that was conducted by Cornish et al., it was reported that preptin has an effect that increases bone mineralization (11).

It was shown in previous studies that preptin activates the ERK/Mitogen-Activated Protein Kinase (MAPK) pathway, and increases the level of Connective Tissue Growth Factor (CTGF) (31). It was also found that there is a correlation between serum preptin and osteocalcin levels in humans (32). It was shown that preptin increases osteogenesis by activating Wnt/ β -catenin signaling

pathway (10). It is known that the same signaling pathway is influential in the growth and development of chondrocyte (33). This signaling pathway has relations with diseases like deformity of skeletons, dwarfism, osteoporosis, degenerative joint disorders and high-bone mass syndrome (34). In human and animal studies, the relation of Wnt pathway genes with hip OA, knee OA, and osteoporosis was reported (35). In another study, it was shown that the preptin levels are low in female osteoporosis and osteopenia patients compared to healthy women (12). Considering the findings of all previous studies, the idea that "the preptin levels are low in knee OA patients compared to healthy subjects" was our hypothesis. As a matter of fact, it was found in our study that preptin levels were lower at a significant level in female OA patients compared to healthy women. In our study, although CRP levels were found to be higher in OA group compared to the Control Group, CRP levels were within normal limits in both groups. It was reported in previous studies that CRP levels increased in OA patients. There are also several studies arguing that CRP levels are associated with OA disease stage (17,36). No correlations were detected between the grade of the disease (Grade 3 and 4) and CRP levels. In our study, the BMI levels were also higher in the OA Group compared to the Control Group. It is already known that BMI is an important risk factor for OA (37).

CONCLUSION

It was found in the present study that preptin peptide that is known to have anabolic effects on bone metabolism is associated with knee OA disease. Further studies are needed to understand the mechanisms of preptin in patients with knee OA.

Competing interests: The authors declare that they have no competing interest.

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REFERENCES

- Hedbom E, Hauselmann HJ. Molecular aspects of pathogenesis in osteoarthritis: the role of inflammation. *Cell Mol Life Sci* 2002;59:45-53.
- Goldring MB. The role of the chondrocyte in osteoarthritis. *Arthritis Rheum* 2000;43:1916-26.
- Min BH, Kim HJ, Lim H, et al. Effects of ageing and arthritic disease on nitric oxide production by human articular chondrocytes. *Exp Mol Med* 2001;33:299-302.
- Hunter DJ. Osteoarthritis. *Best Pract Res Clin Rheumatol* 2011;25:801-14.
- Zhang W, Moskowitz RW, Nuki G, et al. OARSI recommendations for the management of hip and knee osteoarthritis, part I: critical appraisal of existing treatment guidelines and systematic review of current research evidence. *Osteoarthritis Cartilage* 2007;15:981-1000.

6. Zhang W, Moskowitz RW, Nuki G, et al. OARSI recommendations for the management of hip and knee osteoarthritis, Part II: OARSI evidence-based, expert consensus guidelines. *Osteoarthritis Cartilage* 2008;16:137-62.
7. Hunter DJ, Lo GH. The management of osteoarthritis: an overview and call to appropriate conservative treatment. *Rheum Dis Clin North Am* 2008;34:689-712.
8. Kacar C, Gilgil E, Urhan S, et al. The prevalence of symptomatic knee and distal interphalangeal joint osteoarthritis in the urban population of Antalya, Turkey. *Rheumatol Int* 2005;25:201-4.
9. Buchanan CM, Phillips AR, Cooper GJ. Preptin derived from proinsulin-like growth factor II (proIGF-II) is secreted from pancreatic islet beta-cells and enhances insulin secretion. *Biochem J* 2001;360:431-9.
10. Xiao C, Li W, Lu T, et al. Preptin Promotes Proliferation and Osteogenesis of MC3T3-E1 Cells by Upregulating beta-Catenin Expression. *IUBMB Life* 2019.
11. Cornish J, Callon KE, Bava U, et al. Preptin, another peptide product of the pancreatic beta-cell, is osteogenic in vitro and in vivo. *Am J Physiol Endocrinol Metab* 2007;292:E117-22.
12. Li N, Zheng YB, Han J, et al. Lower circulating preptin levels in male patients with osteoporosis are correlated with bone mineral density and bone formation. *BMC Musculoskelet Disord* 2013;14:49.
13. Nazari Soltan Aahmad S, Nourollahi S, Kazerouni F, et al. Investigation of the relation between bone mass density and serum preptin levels in pre- and postmenopausal women. *J Bone Miner Metab* 2018;36:710-5.
14. Altman R, Asch E, Bloch D, et al. Development of criteria for the classification and reporting of osteoarthritis. Classification of osteoarthritis of the knee. Diagnostic and Therapeutic Criteria Committee of the American Rheumatism Association. *Arthritis Rheum* 1986;29:1039-49.
15. Kellgren JH, Lawrence JS. Radiological assessment of osteo-arthrosis. *Ann Rheum Dis* 1957;16:494-502.
16. Massart F, Reginster JY, Brandi ML. Genetics of menopause-associated diseases. *Maturitas* 2001;40:103-16.
17. Nguyen LT, Sharma AR, Chakraborty C, et al. Review of Prospects of Biological Fluid Biomarkers in Osteoarthritis. *Int J Mol Sci* 2017;18.
18. Blom AB, van Lent PL, Libregts S, et al. Crucial role of macrophages in matrix metalloproteinase-mediated cartilage destruction during experimental osteoarthritis: involvement of matrix metalloproteinase 3. *Arthritis Rheum* 2007;56:147-57.
19. Zhen G, Wen C, Jia X, et al. Inhibition of TGF-beta signaling in mesenchymal stem cells of subchondral bone attenuates osteoarthritis. *Nat Med* 2013;19:704-12.
20. Kumm J, Tamm A, Lintrop M. Diagnostic and prognostic value of bone biomarkers in progressive knee osteoarthritis: a 6-year follow-up study in middle-aged subjects. *Osteoarthritis Cartilage* 2013;21:815-22.
21. Lee NK, Sowa H, Hinoi E, et al. Endocrine regulation of energy metabolism by the skeleton. *Cell* 2007;130:456-69.
22. Hylka VW, Teplow DB, Kent SB, et al. Identification of a peptide fragment from the carboxyl-terminal extension region (E-domain) of rat proinsulin-like growth factor-II. *The Journal of biological chemistry* 1985;260:14417-20.
23. Kiess W, Paquette J, Koepf G, et al. Proinsulin-like growth factor-II overexpression does not alter monoallelic H19 gene expression in transfected human embryonic kidney fibroblasts. *Biochem Biophys Res Commun* 1999;255:226-30.
24. Naot D, Cornish J. Cytokines and Hormones That Contribute to the Positive Association between Fat and Bone. *Front Endocrinol (Lausanne)* 2014;5:70.
25. Caplan AL. Mesenchymal stem cells. *J Orthop Res* 1991;9:641-50.
26. Pittenger MF, Mackay AM, Beck SC, et al. Multilineage potential of adult human mesenchymal stem cells. *Science* 1999;284:143-7.
27. Maruotti N, Corrado A, Cantatore FP. Osteoblast role in osteoarthritis pathogenesis. *J Cell Physiol* 2017;232:2957-63.
28. Kwan Tat S, Pelletier JP, Amiable N, et al. Activation of the receptor EphB4 by its specific ligand ephrin B2 in human osteoarthritic subchondral bone osteoblasts. *Arthritis Rheum* 2008;58:3820-30.
29. Kwan Tat S, Pelletier JP, Lajeunesse D, et al. The differential expression of osteoprotegerin (OPG) and receptor activator of nuclear factor kappaB ligand (RANKL) in human osteoarthritic subchondral bone osteoblasts is an indicator of the metabolic state of these disease cells. *Clin Exp Rheumatol* 2008;26:295-304.
30. Mansell JP, Tarlton JF, Bailey AJ. Biochemical evidence for altered subchondral bone collagen metabolism in osteoarthritis of the hip. *Br J Rheumatol* 1997;36:16-9.
31. Liu YS, Lu Y, Liu W, et al. Connective tissue growth factor is a downstream mediator for preptin-induced proliferation and differentiation in human osteoblasts. *Amino Acids* 2010;38:763-9.
32. El-Eshmawy M, Abdel Aal I. Relationships between preptin and osteocalcin in obese, overweight, and normal weight adults. *Appl Physiol Nutr Metab* 2015;40:218-22.
33. Sun Y, Wang F, Sun X, et al. CX3CR1 regulates osteoarthrosis chondrocyte proliferation and apoptosis via Wnt/beta-catenin signaling. *Biomed Pharmacother* 2017;96:1317-23.
34. Usami Y, Gunawardena AT, Iwamoto M, et al. Wnt signaling in cartilage development and diseases: lessons from animal studies. *Lab Invest* 2016;96:186-96.
35. Velasco J, Zarrabeitia MT, Prieto JR, et al. Wnt pathway genes in osteoporosis and osteoarthritis: differential expression and genetic association study. *Osteoporos Int* 2010;21:109-18.
36. Saberi Hosnijeh F, Siebuhr AS, Uitterlinden AG, et al. Association between biomarkers of tissue inflammation and progression of osteoarthritis: evidence from the Rotterdam study cohort. *Arthritis Res Ther* 2016;18:81.
37. Knapik JJ, Pope R, Orr R, et al. Osteoarthritis: pathophysiology, prevalence, risk factors, and exercise for reducing pain and disability. *J Spec Oper Med* 2018;18:94-102.